

A NEW CATALYST FOR THE SYNTHESIS OF NUCLEOSIDES
(ORGANOPHOSPHORUS COMPOUNDS, PART VI)

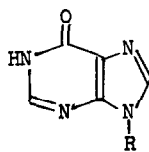
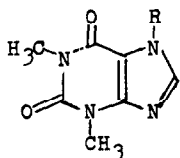
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We wish to report that bis-(p-nitrophenyl) hydrogen phosphate and the methyl derivative act as effective catalysts for the fusion method developed recently for the synthesis of purine nucleosides. The method has involved heating fully O-acetylated D-pentoses and hexoses with various purines in the presence of catalysts like p-toluenesulfonic acid, zinc chloride, concentrated sulfuric acid, sulfamic acid, and chloroacetic acid.

Recently, we investigated the fusion of penta-O-acetyl- β -D-glucopyranose(I) and two purines(II and III) employing bis-(p-nitrophenyl)hydrogen phosphate(VIII) and the related compounds as catalysts.

Bis-(p-nitrophenyl) hydrogen phosphate was used after drying for thirty minutes at 100° under a pressure 1 mm. Hg.



II. R: H

III. R: H

IV. R: 2',3',4',6'-tetra-O-acetyl- β -D-glucopyranosyl residueV. R: 2',3',4',6'-tetra-O-acetyl- β -D-glucopyranosyl residueVI. R: β -D-glucopyranosyl residueVII. R: β -D-glucopyranosyl residue

When theophylline(II) was fused with I in the presence of VIII at 140°-160° for one hour, N⁷-(2',3',4',6'-tetra-O-acetyl- β -D-glucopyranosyl)theophylline(IV) was obtained in 47% yield. m.p. 145°-146°.

Anal. Calcd. for C₂₁H₂₆O₁₁N₄: C, 49.41; H, 5.13; N, 10.97.

Found: C, 49.38; H, 5.16; N, 10.95.

Removal of the acetyl groups from IV in methanolic ammonia afforded N⁷-(β -D-glucopyranosyl) theophylline(VI) in 75% yield. Recrystallization from aqueous methanol gave colorless needles. m.p. 266°-268°.

Anal. Calcd. for C₁₈H₁₈O₇N₄: C, 45.61; H, 5.30; N, 16.37.

Found: C, 45.58; H, 5.29; N, 16.23.

$\lambda_{max}^{H_2O}$ 275 μ . (E 8800), $[\alpha]_D^{19}$ -3.56°(c, 1.49, water).

(Reported: m.p. 267-269°, ⁽⁸⁾ 272°, ⁽⁹⁾ $[\alpha]_D$ -2.33°(water), ⁽⁹⁾ $[\alpha]_D^{19}$ +39.09°(c, 0.92, water). ⁽¹⁰⁾)

The formation of the glycoside link at position 7 of theophylline was supported by the ultraviolet absorption spectra of the compound VI, N⁷-methyltheophylline (caffeine, $\lambda_{\text{max}}^{\text{pH } 5}$ 273 m μ .), and N⁹-methyltheophylline (isocaffeine, $\lambda_{\text{max}}^{\text{pH } 5}$ 239 and 267 m μ .).⁽¹¹⁾ The anomeric configuration was assigned as β - from the coupling constant for the C₁ proton of glucose (FIG. I., J_{1,2}=8.7-8.8 c.p.s.).^(12,13) The infrared spectra of the compound VI and N⁷-(β -D-glucopyranosyl)theophylline synthesized according to the Fischer-Helferich procedure were superimposable. Further, the optical rotatory dispersion curve⁽¹⁴⁾ was negative plain as shown in FIG.II.

Similarly, the fusion of II with I employing bis-(p-nitrophenyl) methylphosphate(IX) instead of VIII afforded IV in 31% yield, whereas the desired product(IV) could not be obtained when tris-(p-nitrophenyl) phosphate was used and unreacted theophylline was recovered almost quantitatively.

For the comparison of the catalytic effects of these organophosphorus compounds with that of previously known catalysts, p-toluenesulfonic acid and ethyl polyphosphate^(15,16,17) were used in similar manner described above. But the yield of IV were 20% employing p-toluenesulfonic acid and 31% when ethyl polyphosphate was used.⁽³⁾ Moreover, earlier paper reported a few percent yield in the synthesis of IV using zinc chloride. Thus, it is apparent that the catalytic effect of bis-(p-nitrophenyl) hydrogen phosphate is superior than known catalysts for fusion procedure. Considerable activity of bis-(p-nitrophenyl) methylphosphate may be explained by the demethylation during the fusion.

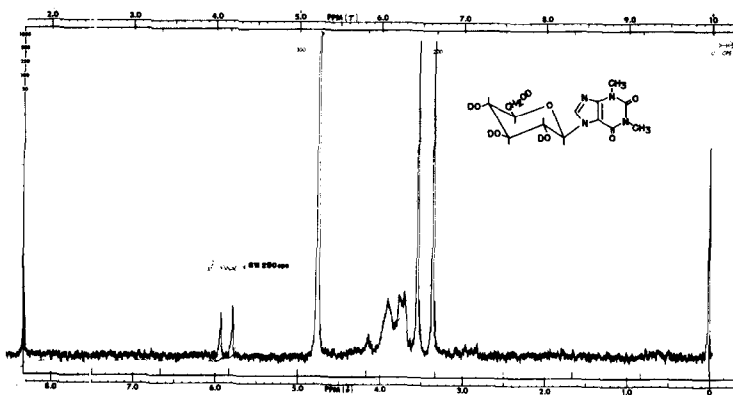


FIG. I. N.m.r. spectrum of VI measured in D_2O .

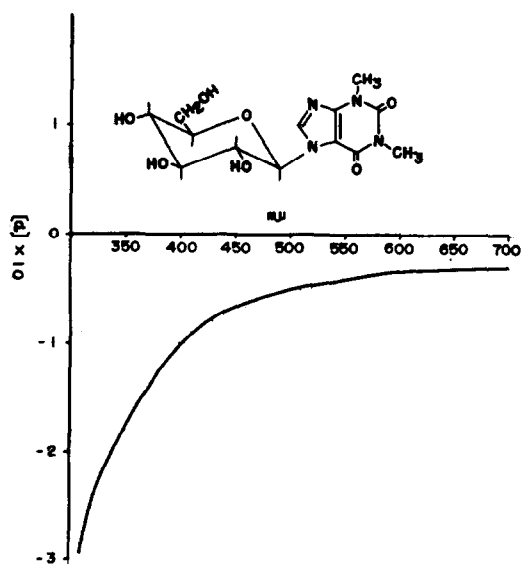


FIG. II. Optical rotatory dispersion curve of VI.

In addition, the fusion of N¹-acetyl hypoxanthine* and I in the presence of VIII at 150°-160° for thirty minutes gave N⁹-(2',3',4',6'-tetra-O-acetyl-β-D-glucopyranosyl)hypoxanthine (V) in 13% yield after recrystallization from ethyl acetate. m.p. 276°-277°.

Anal. Calcd. for C₁₉H₂₂O₁₀N₄: C, 48.93; H, 4.75; N, 12.01.

Found: C, 48.92; H, 4.85; N, 12.01.

$\lambda_{\text{max.}}^{\text{abs. EtOH}}$ 244 mμ. (13,300), $[\alpha]_{\text{D}}^{19}$ -26.4° (c, 0.73, chloroform).
(18)
(Reported: m.p. 265-267.5°, $[\alpha]_{\text{D}}^{28}$ -19.2° (c, 3.06, chloroform).

Removal of the acetyl groups from V in methanolic ammonia gave N⁹-(β-D-glucopyranosyl) hypoxanthine(VII) in 78% yield.

m.p. 272°-273° (from 50% aqueous ethanol).

Anal. Calcd. for C₁₁H₁₄O₆N₄.H₂O: C, 41.77; H, 5.10; N, 17.72.

Found: C, 41.70; H, 5.15; N, 17.64.

$\lambda_{\text{max.}}^{\text{H}_2\text{O}}$ 248.5 mμ. (ε 12,600), $\lambda_{\text{max.}}^{0.05\text{N-HCl}}$ 248.5 mμ. (ε 12,100),
 $\lambda_{\text{max.}}^{0.05\text{N-NaOH}}$ 253.5 mμ. (ε 13,700), $[\alpha]_{\text{D}}^{20}$ -38.4° (c, 2.11, N-NaOH).

The allocation of C₁ of glucose to position 9 of hypoxanthine was based on the ultraviolet spectra of the compound obtained, authentic N⁹-(VII)**N⁷-(β-D-glucopyranosyl) hypoxanthine ($\lambda_{\text{max.}}^{0.05\text{N-HCl}}$ 254 mμ., $\lambda_{\text{max.}}^{0.05\text{N-NaOH}}$ 263 mμ.)**N⁹-, and N⁷-methylhypoxanthine. The assignment of the β-configuration at C₁ of

*

This derivative was prepared in refluxing hypoxanthine with acetic anhydride and N,N-dimethylformamide. m.p. 238°-240°. The position of the acetyl group linked was inferred from the infrared spectra.

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Kindly given to us by Mr. H. Yamazaki who prepared both N⁹-, and N⁷-(β-D-glucopyranosyl) hypoxanthine by Davoll-Low method.

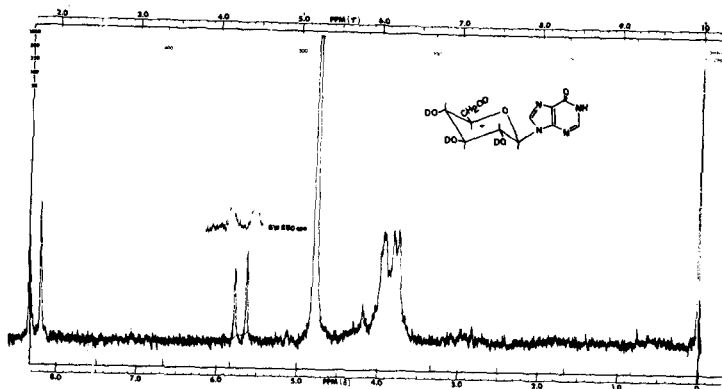


FIG. III. N.m.r. spectrum of VII measured in D_2O .

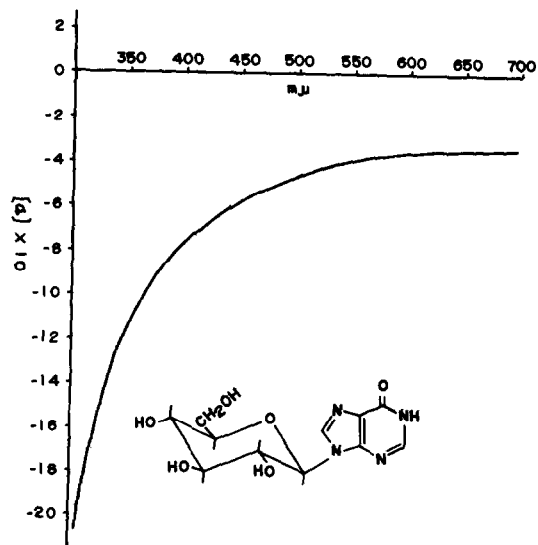


FIG. IV. Optical rotatory dispersion curve of VII.

the compound was based on the coupling constant, $J_{1,2}$ of the proton at C_1 , and the equation of Karplus. (FIG.III., $J_{1,2}^{(20)} = 8.7-8.8$ c.p.s.). Moreover, upon admixture with an authentic specimen, the compound showed no depression of melting point. Further, the optical rotatory dispersion curve is shown in FIG.IV. The o.r.d. curves were measured on a JASCO automatic recording spectropolarimeter and the n.m.r. spectra were recorded on a Varian A-60 spectrometer.

Work with other bases and sugars in this fusion procedure using VIII is in progress and an improved synthesis of nebularine, a nucleoside antibiotics, will be reported independently. We are grateful to Dr. T. Mitsui for performing the microanalyses and Dr. T. Okuda for measuring the o.r.d. curves.

REFERENCES

- (1) T. Simadate, Y. Ishido, and T. Sato, Nippon Kagaku Zasshi, 82, 938(1961).
- (2) T. Simadate, ibid., 82, 1268, 1270 (1961).
- (3) T. Simadate, ibid., 83, 212 (1962).
- (4) Y. Ishido, and T.Sato, Bull. Chem. Soc. Japan, 34, 1347(1961).
- (5) Y. Ishido, A. Hosono, S. Isome, A. Maruyama, and T. Sato, ibid., 37, 1389 (1964).
- (6) M. J. Robins, W. A. Bowles, and R. K. Robins, J. Amer. Chem. Soc. 86, 1251 (1964).
- (7) T. Hashizume and H. Hiro, Japanese Pat. 437,638 (Jan. 18, 1965).
- (8) J. M. Gulland, and T. F. Macrae, J. Chem. Soc., 1933, 662.
- (9) E. Fischer, and B. Helferich, Ber., 47, 210 (1914).

- (10) K. Onodera, and H. Fukumi, Agr. Biol. Chem. 27, 864 (1963).
- (11) J. M. Gulland, E. R. Holiday, and T. F. Macrae,
J. Chem. Soc., 1974, 1639.
- (12) R. U. Lemieux, R. K. Kullnig, H. J. Bernstein and
W. G. Schneider, J. Amer. Chem. Soc., 80, 6098 (1958).
- (13) R. U. Lemieux and J. W. Lown, Can. J. Chem., 41, 889 (1963).
- (14) I. Listowsky, G. Avigad, and S. England,
J. Amer. Chem. Soc., 87, 1765 (1965).
- (15) G. Schramm, Angew. Chem., 74, 53 (1962)
- (16) W. Pollmann and G. Schramm, Biochim. Biophys. Acta,
80, 1 (1964).
- (17) G. Burkhardt, M. P. Klein, and M. Calvin,
J. Amer. Chem. Soc., 87, 591 (1965).
- (18) T. Nishimura and B. Shimizu, Agr. Biol. Chem., 28, 224 (1964).
- (19) J. M. Gulland and E. R. Holiday, J. Chem. Soc., 1936, 765.
- (20) M. Karplus, J. Chem. Phys., 30, 11 (1959).